



# The Valley Ponds



## Stage 1 Germination Report

### Introduction

This report for the ACT Urban Waterways Coordinator, Edwina Robinson, outlines Stage 1 of Greening Australia's rehabilitation work at The Valley Ponds.

Stage 1 involved identifying six zones for recovering wetland plants, relocating representative plant material to a small dam on site, and collecting seed cores (mud samples) from three zones for germinating at Greening Australia's nursery in Aranda.

The aim of Stage 1 works was to conserve locally adapted genetic material, for reintroduction to the

Valley Ponds in mid 2012 following completion of the constructed wetlands.

### Materials and Methods

#### Plant material

Six zones from which to recover plant material were identified in and around the main dam, ensuring submerged and emergent plants were recovered.

61 styrofoam boxes were filled with plant material from the main dam in August 2011 (see Figure 1) and relocated to the small dam in the south-western section of the Valley Ponds. Holes were cut into the bottom and sides of the boxes to ensure the roots of the plants remained moist. The boxes were initially placed in the shallow (emergent) zone of the dam.

The main genera recovered included *Juncus*, *Eleocharis*, and *Myriophyllum*. Particular species targeted for collection included *Amphibromus nervosus* and *Ranunculus papulentus*.

Ten buckets full of *Myriophyllum crispatum* were relocated to the small dam. Unlike the other plant material that was kept in the



Figure 1 - collecting plant material



styrofoam boxes, the *Myriophyllum* as placed directly into the dam. The request to relocate *Potamogeton tricarlinatus* to the small dam could not be fulfilled as the species was not found in the main dam.

A dry spell in January 2012 led to a marked drop in the water level of the small dam. The styrofoam boxes were therefore moved further into the dam to ensure plant roots remained moist (see Figure 2).



**Figure 2 – Plant material in the small dam**

### Seed cores

The following methodology was adapted from a wetland trial conducted by Dr Patricia Murray in 2004 (Murray 2008).

Seed cores (mud samples) were collected from the emergent/littoral zone (or high water mark), submergent zone (or mid water mark) and the open water zone (or low water mark), at both the large dam and the nearby frog pond (smaller dam near the Scout hall) in September 2011. The mud was collected using a shovel and buckets (See Figure 3). A total of 24 buckets' worth of seed cores was collected from the large dam, and 12 buckets collected from the frog pond.



**Figure 3 – Collecting seed cores**

Half the seed cores from each of the three collection zones (high water, mid water and open water) were dried at ambient temperature, and half were kept moist, to help indicate whether a trigger event like wetting improves the rate of germination.

The seed cores (both dried and moist) were placed in 2L ice-cream containers on top of a mixture of washed sand and vermiculite. Holes were drilled in the bottom of each ice-cream container. The cores were further divided into moist and submerged treatments. Each pair of treatments was placed in a styrofoam box. Submerged treatments were placed on the bottom of the box, while the moist treatments were raised above the bottom using an inverted ice-cream container. The styrofoam boxes were filled with water to a level 2cm above the submerged treatments, which also ensured the moist treatments were sitting in 2cm of water.

The water level in the styrofoam boxes was checked regularly and topped to initial levels when necessary, and the water was changed periodically to prevent algal build-up.

Some dried seed cores were put aside as a source of micro-crustaceans and seed, to be reintroduced later with the plants.

### Results

See tables opposite.

### Discussion

#### Plant material

The relocated plant material has grown well in the styrofoam boxes. The targeted nature of the plant material has resulted in less exotic species than have germinated in the seed cores, with 9 native species and 5 exotic species identified. The main challenge was to keep the roots moist, with the fluctuating water level in the dam.

Weeds will be removed in preparation for planting. At this stage species that are easier to

separate such as *Juncus* will be separated, whereas smaller or more fragile species will be planted in clumps. Care will be taken in separating plants to ensure root damage is minimised.

### Seed cores

Species that germinated included 9 natives and 10 exotics. Comparing the different treatments, a greater abundance and diversity of species was found in the moist than the submerged treatments. There was little difference in abundance between dried and wet samples. This suggests the mud samples did not require drying to trigger germination. There was also no significant difference between samples taken from high, mid and low water marks, with perhaps a few more stems found in the high and mid water samples.

Challenges with the seed cores included algal growth, which necessitated periodic changing of the water. The styrofoam boxes were also exposed to the elements, resulting in occasional inundation during heavy rainfall.

### Next steps

Plants will be monitored until the time of reintroduction to the Valley Ponds. Following the planting, a final report will be produced.

### References

- Brock M (1997) 'Are there seeds in your wetland? Assessing wetland vegetation', Land & Water Resources Research & Development Corporation, University of New England, Department of Land & Water Conservation and Environment Australia, Canberra
- Clifton, E (2004) Murrumbidgee Wetland Seedbank Research Project, Preliminary Report, Land & Water Australia, Canberra
- Murray P (2008) Survey of soil seed bank of Murrumbidgee floodplain wetlands, Land & Water Australia, Canberra

## Results

**Table 1 - Plant material species**

The most abundant native species were *Juncus usitatus*, *Eleocharis acuta*, and *Myriophyllum crispatum*. Notable species were *Amphibromus nervosus* and *Ranunculus papulentus*.

Scientific name	Common name	Origin	Life cycle	No. of stems (~)
<i>Amphibromus nervosus</i>	Common Swamp Wallaby Grass	N	P	<10
<i>Crassula helmsii</i>	Water Stonecrop	N	P	<20
<i>Eleocharis acuta</i>	Common Spikerush	N	P	1000+
<i>Epilobium</i> sp.	Willow Herb	N	P	25
<i>Hydrocotyle peduncularis</i>	Stinking Pennywort	N	P	50+
<i>Juncus articulatus</i>	Jointed Rush	N	P	120
<i>Juncus bufonius</i>	Toad Rush	N	A	100
<i>Juncus usitatus</i>	Common Rush	N	P	100+
<i>Lythrum hyssopifolia</i>	Hyssop Loosestrife	N	A	20+
<i>Myosotis caespitosa</i>	Forget-me-not	E	A	250+
<i>Myriophyllum crispatum</i>	Upright Water Milfoil	N	P	750+
<i>Paspalum dilatatum</i>	Paspalum	E	P	10
<i>Polypogon monspeliensis</i>	Beard Grass	E	P	30
<i>Ranunculus papulentus</i>	Large River Buttercup	N	P	350
<i>Rumex crispus</i>	Dock	E	P	30
<i>Trifolium</i> sp.	Clover	E	P	<10
<i>Veronica anagallis-aquatica</i>	Blue Water Speedwell	E	P	20

(N = native, E = exotic, P = perennial, A = annual)

**Table 2 - Seed core species**

The most abundant native species, *Elatine gratioloides* and *Juncus bufonius*, are both annuals.

Scientific name	Common name	Origin	Life cycle	No. of stems (~)
<i>Amphibromus nervosus</i>	Common Swamp Wallaby Grass	N	P	10
<i>Cirsium vulgare</i>	Spear Thistle	E	A	<5
<i>Cyperus eragrostis</i>	Umbrella Sedge	E	P	75+
<i>Echinochloa crus-galli</i>	Barnyard Grass	E	A	<5
<i>Elatine gratioloides</i>	Waterwort	N	A	500+
<i>Eleocharis acuta</i>	Common Spikerush	N	P	250+
<i>Holcus lanatus</i>	Yorkshire Fog	E	P	10
<i>Juncus articulatus</i>	Jointed Rush	E	P	500+
<i>Juncus bufonius</i>	Toad Rush	N	A	500+
<i>Lythrum hyssopifolia</i>	Hyssop Loosestrife	N	A	200+
<i>Myosotis caespitosa</i>	Forget-me-not	E	A	25+
<i>Myriophyllum crispatum</i>	Upright Water Milfoil	N	P	375+
<i>Ranunculus papulentus</i>	Large River Buttercup	N	P	10
<i>Rumex crispus</i>	Dock	E	P	<5
<i>Trifolium</i> sp.	Clover	E	P	<5
<i>Veronica anagallis-aquatica</i>	Blue Water Speedwell	E	P	10